



# The role of cannabinoids in modulating emotional and non-emotional memory processes in the hippocampus

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Cannabinoid agonists generally have a disruptive effect on memory, learning, and operant behavior that is considered to be hippocampus-dependent. Nevertheless, under certain conditions, cannabinoid receptor activation may facilitate neuronal learning processes. For example, CB<sub>1</sub> receptors are essential for the extinction of conditioned fear associations, indicating an important role for this receptor in neuronal emotional learning and memory. This review examines the diverse effects of cannabinoids on hippocampal memory and plasticity. It shows how the effects of cannabinoid receptor activation may vary depending on the route of administration, the nature of the task (aversive or not), and whether it involves emotional memory formation (e.g., conditioned fear and extinction learning) or non-emotional memory formation (e.g., spatial learning). It also examines the memory stage under investigation (acquisition, consolidation, retrieval, extinction), and the brain areas involved. Differences between the effects of exogenous and endogenous agonists are also discussed. The apparently biphasic effects of cannabinoids on anxiety is noted as this implies that the effects of cannabinoid receptor agonists on hippocampal learning and memory may be attributable to a general modulation of anxiety or stress levels and not to memory *per se*. The review concludes that cannabinoids have diverse effects on hippocampal memory and plasticity that cannot be categorized simply into an impairing or an enhancing effect. A better understanding of the involvement of cannabinoids in memory processes will help determine whether the benefits of the clinical use of cannabinoids outweigh the risks of possible memory impairments.

**Keywords:** cannabinoids, CB<sub>1</sub> receptors, hippocampus, LTP, stress, emotional memory, anxiety, extinction

## INTRODUCTION

Considerable evidence suggests that cannabinoids impair hippocampal-dependent learning and memory processes, such as spatial learning and context-related memory tasks (Sullivan, 2000; Riedel and Davies, 2005). In this review, I will provide evidence that suggests that the effects of cannabinoids on memory and plasticity are complex and depend on several factors, such as the nature of the task (emotional or non-emotional), the memory stage investigated (acquisition, retrieval, and extinction), and the experimental model used. Naturally, the behavioral effects of cannabinoids on memory may vary as a function of dose, route of administration, and the specific drug used.

## CANNABINOID RECEPTORS IN THE HIPPOCAMPUS

Cannabis has a long history of consumption both for recreational and medicinal uses. The main psychoactive constituent of marijuana, delta-9-tetrahydrocannabinol (THC), was identified in 1964 (Gaoni and Mechoulam, 1964) and this discovery led to the identification of the endogenous endocannabinoid (eCB) system. This system includes cannabinoid receptors (CB<sub>1</sub> and CB<sub>2</sub>), eCBs [anandamide and 2-arachidonoyl-glycerol (2-AG)], enzymes involved in their synthesis and metabolism [fatty acid amide hydrolase (FAAH) for anandamide and the monoacylglycerol lipase (MAGL) for 2-AG], and an eCB transporter (Devane et al., 1992; Freund et al., 2003; Kogan and Mechoulam, 2006). Recent cDNA cloning of the key enzymes such as *N*-acylphosphatidylethanolamine-hydrolyzing

phospholipase D (NAPE-PLD) and diacylglycerol lipase (DAGL) accelerated molecular biological studies on the eCB biosyntheses (Bisogno et al., 2003; Okamoto et al., 2004). eCBs are synthesized “on demand” at the post-synaptic sites of neurons after an increase in neural activity and calcium ion influx, and are then released into the synaptic cleft. Their main function appears to be the suppression of neurotransmitter release from the presynapse. Thus, eCBs act as retrograde neurotransmitters, modulating other neurotransmitter systems.

CB<sub>1</sub> and CB<sub>2</sub> are metabotropic receptors coupled to G-proteins of the Gi/o type. CB<sub>1</sub> receptors are localized mainly in the central nervous system, but are also present in a variety of peripheral tissues; they are among the most abundant and widely distributed G-protein coupled receptors in the brain. CB<sub>1</sub> receptors are expressed in multiple brain areas, including the olfactory bulb, neocortex, pyriform cortex, hippocampus, amygdala, basal ganglia, thalamic and hypothalamic nuclei, cerebellar cortex, and brainstem nuclei (Herkenham et al., 1990, 1991; Katona et al., 2001). CB<sub>2</sub> receptors are mostly peripherally located on immunological tissues, but they have also been found within the central nervous system on neurons and glial cells with their expression mainly related to conditions of inflammation (Galiegue et al., 1995; Schat et al., 1997; Begg et al., 2005). More recent immunohistochemical analyses have revealed the presence of CB<sub>2</sub> receptors in apparently neuronal and glial processes in diverse rat brain areas, including the cerebellum and hippocampus (Van Sickle et al., 2005; Onaivi et al., 2006).

In the hippocampus, CB<sub>1</sub> receptors are expressed at an especially high density in the dentate gyrus, CA1, and CA3 regions (Herkenham et al., 1990, 1991; Matsuda et al., 1990; Tsou et al., 1998). CB<sub>1</sub> receptors are predominantly localized on the axon terminals and preterminal segments of cholecystokinin (CCK)-expressing GABAergic interneurons (Nyíri et al., 2005); however, they have also been demonstrated to inhibit glutamatergic transmission in cultured hippocampal cells (Shen, et al., 1996). CB<sub>1</sub> receptors located on GABAergic axon terminals are activated by lower concentrations of cannabinoid receptor agonists than CB<sub>1</sub> receptors located on glutamatergic terminals (Ohno-Shosaku et al., 2001; Hoffman et al., 2007) and CB<sub>1</sub> receptor expression is significantly lower on glutamatergic terminals than on GABA axon terminals in the hippocampus (Katona et al., 2006; Kawamura et al., 2006). Specifically, activation of hippocampal CB<sub>1</sub> receptors decreases GABA release (Katona et al., 1999; Hajos et al., 2000; Hoffman and Lupica, 2000; Hoffman et al., 2003). The CB<sub>1</sub>-containing GABAergic interneurons are thought to control oscillatory electrical activity in the hippocampus in the theta and gamma frequencies, which plays a role in synchronizing pyramidal cell activity (Hoffman and Lupica, 2000).

Overall, the evidence favors a predominant role for GABAergic pathways in the effects of cannabinoids on hippocampal-dependent memory processes.

### CANNABINOID AGONISTS IMPAIR HIPPOCAMPAL-DEPENDENT LEARNING AND MEMORY

In humans, non-human primates, and rodents, cannabinoids impair the performance of a wide variety of memory tasks that share the common feature of requiring the hippocampus for normal performance (Sullivan, 2000; Davies et al., 2002; Riedel and Davies, 2005). In laboratory rodents, activation of cannabinoid receptors via THC or synthetic analogues such as WIN 55,212-2, CP55940, HU-210 or the endogenous agonist anandamide impairs learning (Davies et al., 2002). Administration of THC disrupts hippocampal-dependent learned behavior in operant and spatial maze models of memory (Nakamura et al., 1991; Heyser et al., 1993; Lichtman et al., 1995; Brodtkin and Moerschbaecher, 1997; Mallet and Beninger, 1998; Ferrari et al., 1999; Varvel et al., 2001). For example, systemic THC administration (2–6 mg/kg i.p.) impairs working memory tested in the radial-arm spatial task and the cannabinoid antagonist SR141716A (1–10 mg/kg) prevents these deficits in a dose-dependent manner (Lichtman and Martin, 1996). Similarly, THC (8 mg/kg) impairs the acquisition of spatial learning in the water maze and the performance of mice in a working memory task, while consolidation and retrieval of a previously learned task are not affected. Pre-treatment with the antagonist SR 141716A (1 mg/kg i.p.) prevents these learning deficits (Da and

Takahashi, 2002). Additionally, systemic administration of THC or the synthetic cannabinoid receptor agonist WIN 55,212-2 reliably impairs performance in delayed-match-to-sample and delayed-non-match-to-sample tasks, and this is accompanied by decreases in hippocampal cell firing during the sample phases of the task (Heyser et al., 1993; Hampson and Deadwyler, 1999, 2000).

Overall, the literature discussed above suggests that activation of cannabinoid receptors impairs learning. However, since the agonists were systemically infused, most of these experiments do not specifically show that cannabinoids impair learning and memory via action on the hippocampus. Rather, the involvement of the hippocampus is assumed because it is an important target for systemically administered cannabinoids and because most of the paradigms described are spatial tasks known to be hippocampus-dependent.

More recent research has directly tested whether specific administration of cannabinoids into the hippocampus would have similar effects (summarized in **Table 1**). Intrahippocampal infusions of the agonists CP55940, THC, or WIN 55,212-2 were found to disrupt performance in the radial-arm maze, and in T-maze delayed alternation, passive avoidance, spatial learning, and place recognition memory tasks (Lichtman et al., 1995; Mishima et al., 2001; Egashira et al., 2002; Suenaga and Ichitani, 2008; Suenaga et al., 2008; Wegener et al., 2008; Abush and Akirav, 2010). For example, activation of hippocampal cannabinoid receptors by the agonist WIN 55,212-2 (1–2 µg) dose-dependently decreases the exploration of an object in a new place, and this effect is antagonized by pre-treatment with the cannabinoid receptor antagonist AM 281 (2 mg/kg, i.p.; Suenaga and Ichitani, 2008). WIN 55,212-2 (5 µg) injected into the dorsal hippocampus increases the number of reference memory errors in the eight-arm radial-maze task, suggesting impairment of memory retrieval (Wegener et al., 2008). Additionally, post-training intrahippocampal administration of WIN 55,212-2 (2.5 and 5 µg) disrupts long-term spatial memory, but not acquisition or short-term memory, in a rat reference memory task in the water maze (Yim et al., 2008). We have recently found that WIN 55,212-2 administered systemically (0.5 mg/kg) or specifically into the hippocampal CA1 area (5 µg/side) before massed training in the Morris water maze impairs spatial learning (Abush and Akirav, 2010). Thus experiments that specifically targeted the hippocampus confirm the implications of the earlier systemic research as to the impairing effect of cannabinoids on hippocampal-dependent learning and memory.

### CANNABINOID AGONISTS IMPAIR HIPPOCAMPAL SYNAPTIC PLASTICITY

In neuronal circuits, memory storage depends on activity-dependent modifications in synaptic efficacy, such as long-term potentiation (LTP) and long-term depression (LTD), which are

**Table 1 | Effects of intra-dorsal hippocampal WIN 55,212-2 on learning and memory.**

Doses (µg)	Task	Memory stage	Effects	References
1–2	Place recognition	Short-term retrieval	Impair	Suenaga and Ichitani (2008)
5	Radial-maze	Long-term retrieval	Impair	Wegener et al. (2008)
2.5 and 5	Spatial (water maze)	Long-term retrieval	Impair	Yim et al. (2008)
5	Spatial (water maze)	Acquisition	Impair	Abush and Akirav (2010)

the two main forms of synaptic plasticity in the brain. A key feature of LTP and LTD is that a short period of synaptic activity (either high- or low-frequency stimulation) can trigger persistent changes in synaptic transmission lasting at least several hours and often longer. This single property initially led investigators to suggest that these forms of plasticity are the cellular correlate of learning (Bliss and Gardner-Medwin, 1973; Bliss and Lomo, 1973). Indeed, efforts to understand synaptic plasticity are driven by the belief that such synaptic modifications might occur during learning and memory. However, it is extremely difficult to demonstrate directly that learning-induced synaptic changes occur following experience.

The mechanisms underlying synaptic plasticity have been studied more intensely in the hippocampus than in any other brain region. Both forms of synaptic plasticity have been studied most intensively at the Schaffer collateral–CA1 synapses of the hippocampus because of the established role of the CA1 area in spatial memory (Behr et al., 2009). LTP and LTD are thought to be involved in memory formation at glutamatergic synapses in the hippocampus. Cannabinoids appear to work by reducing glutamate release below the level needed to activate *N*-Methyl-D-aspartate (NMDA) receptors that are required for LTP and LTD (Shen et al., 1996; Misner and Sullivan, 1999). CB<sub>1</sub> receptors are capable of regulating both inhibitory and excitatory neurotransmitter release in the hippocampus and are thus capable of exerting subtle control over synaptic plasticity.

Most of our knowledge about cannabinoids and activity-dependent changes in synaptic strength comes from studies performed at excitatory synapses, largely using acute hippocampal slices as the experimental model (Chevalyere et al., 2006). Cannabinoid receptor activation inhibits both LTP and LTD induction in the hippocampal slice. The inhibition of LTP in field potentials in the CA1 region has been demonstrated using THC, HU-210, WIN 55,212-2, 2-AG, and anandamide (Nowicky et al., 1987; Collins et al., 1994, 1995; Terranova et al., 1995; Misner and Sullivan, 1999) and has been found recently to inhibit hippocampal LTD of CA1 field potentials as well (Misner and Sullivan, 1999). The impairment in the induction of LTP in the CA1 is blocked by cannabinoid antagonists such as SR141716A.

We have recently examined cannabinoid modulation of LTP and LTD in a different experimental model: acute anesthetized rats. Using this experimental condition, we found that i.p. administration of WIN 55,212-2 or the CB<sub>1</sub> receptor antagonist AM251 at the doses tested impairs LTP in the Schaffer collateral–CA1 projection, with no effect on LTD (Abush and Akirav, 2010; see **Figure 1**).

de Oliveira Alvares et al. (2006) have also demonstrated impairment of LTP in a CA1 slice preparation following AM251 administration. Sokal et al. (2008) found that the CB<sub>1</sub> receptor antagonist SR141716A blocked the potentiation of the fEPSP slope observed following HFS to the perforant path. However, other studies conducted on hippocampal slices of the Schaffer collateral–CA1 synapses have shown that CB<sub>1</sub> blockade favors LTP in the hippocampus (Slanina et al., 2005) and that mice lacking CB<sub>1</sub> receptors show enhanced LTP (Bohme et al., 2000). However, in the study by Slanina et al. (2005), the drug was present throughout the experiment and LTP was elicited by moderate stimulations (20 or 50 pulses). Thus, the discrepancies with our findings could result

from the examination of field potential in an intact rat model versus slices, or from various methodological issues, such as different stimulation protocols, different drug doses, etc.

## EFFECTS OF CANNABINOID AGONISTS ON EMOTIONAL AND NON-EMOTIONAL MEMORY

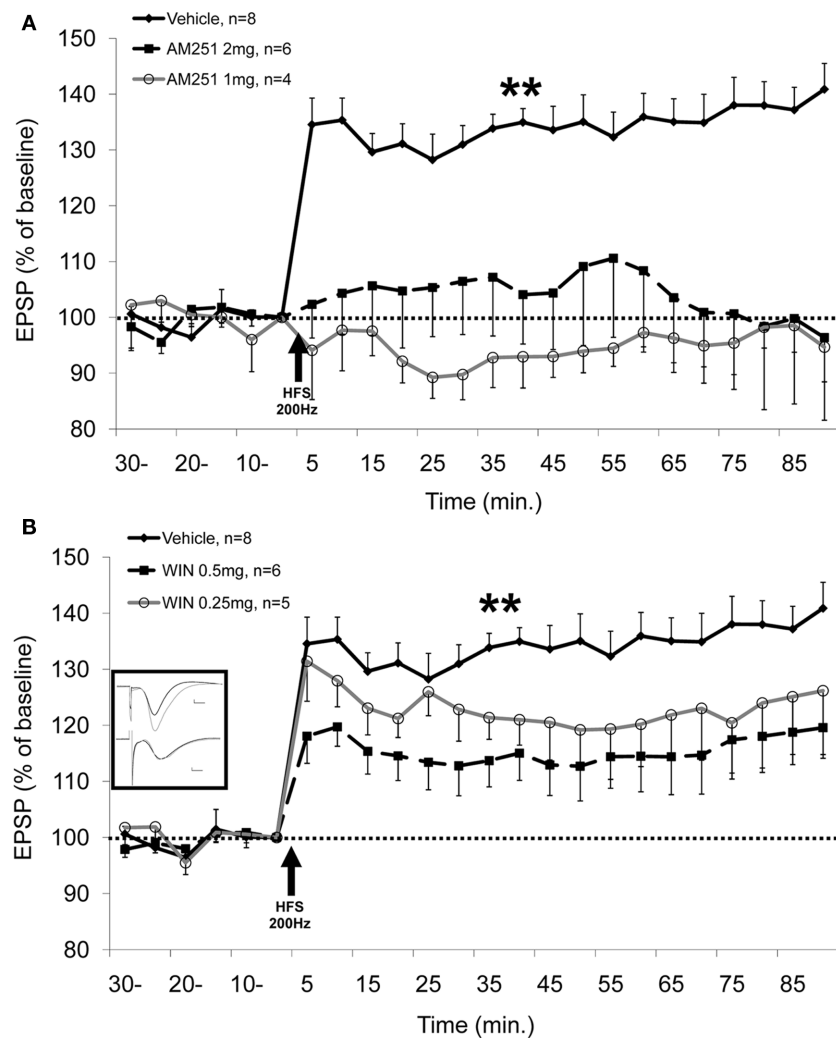
Although considerable evidence suggests that activation of CB<sub>1</sub> receptors can induce learning and memory impairments (Sullivan, 2000; Robinson et al., 2003; O'Shea et al., 2004; Varvel et al., 2005), CB<sub>1</sub> receptors are essential for the extinction of conditioned fear associations (Marsicano et al., 2002), indicating an important role for this receptor in neuronal emotional learning and memory.

## ROLE OF THE CANNABINOID SYSTEM IN EXTINCTION

Extinction was established as a tool to treat conditioned fear by Freud in the 1920s. It has become widely accepted that a deficit in the capacity to extinguish memories of fear is at the root of fear disorders as a result of the distinction between those who do and do not develop serious symptoms after fearsome experiences, and the fact that fear disorders are treated with therapy based on extinction procedures. Moreover, panic attacks, phobias, and particularly post-traumatic stress disorder (PTSD) are viewed by many as a deficit of extinction that should therefore be treated by an intensification of extinction (Charney et al., 1993; Wessa and Flor, 2007; Milad et al., 2008).

Conditioned fear is induced by pairing a neutral, conditioned stimulus (CS; e.g., a light, a tone, or a context) with an aversive stimulus (unconditioned stimulus, US; e.g., a mild footshock) that evokes a measurable fear response. Experimental extinction learning occurs when a CS that previously predicted a US no longer does so, and over time, the conditioned response (e.g., freezing or elevated skin conductance responses) decreases. Extinction learning involves the ventromedial prefrontal cortex (PFC), amygdala, and hippocampus (Milad and Quirk, 2002; Phelps et al., 2004; Bouton et al., 2006). PTSD patients continue to re-experience the traumatic event over a long timeframe and avoid trauma-related stimuli, even though they recognize that the traumatic event is no longer occurring. It has been suggested that dysfunctional fear extinction plays an important role in the development of clinical symptoms, such as reexperiencing trauma in PTSD (Rothbaum and Davis, 2003; Milad et al., 2006; Quirk et al., 2006; Rauch et al., 2006). PTSD patients also demonstrate impaired extinction in the aftermath of new trauma. For example, Milad et al. (2008) have shown deficient extinction recall as measured in skin conductance response in a 2-day fear conditioning and extinction procedure in PTSD patients.

Clearly, animal models do not entirely mimic the complex features of psychiatric disorders. However, they can predict the clinical effects of substances and provide insights into the biological mechanisms of these diseases. Marsicano et al. (2002) found that CB<sub>1</sub> receptor-deficient mice show normal acquisition and consolidation in a fear conditioning task, but fear extinction is strongly impaired. Impaired extinction is also observed when the antagonist SR141716 is injected systemically into wild-type mice before the extinction trial, indicating that CB<sub>1</sub> receptors are required at the moment of the extinction training. The findings that CB<sub>1</sub> knockout mice exhibit impaired short- and long-term extinction



**FIGURE 1 | CB<sub>1</sub> receptor antagonist and agonist impair the induction of LTP (A)** AM251 injected i.p. (1 or 2 mg/kg) 30 min before application of high frequency stimulation (HFS; 200 Hz) to the Schaffer collateral significantly impairs the induction of LTP in the CA1 compared with the vehicle group ( $P < 0.01$ , vehicle differs from all the groups). No significant difference is observed between the groups before HFS. **(B)** WIN 55,212-2 (0.5 mg/kg) injected i.p. 20 min before application of HFS (200 Hz)

to the Schaffer collateral significantly impairs the induction of LTP in the CA1 compared with the vehicle group ( $P < 0.01$ ). No significant difference is observed between the groups before HFS. Inset: representative traces in the CA1 for vehicle (upper traces) and WIN 0.5 mg (lower traces) groups taken before (black) and 90 min after (gray) HFS to the Schaffer collateral (calibration: 0.2 mV, 10  $\mu$ s). Data published by Abush and Akirav (2010) in *Hippocampus*.

of cue-induced conditioned fear responses have been replicated by other groups for the extinction of both cue- and context-induced fear responses (Finn et al., 2004; Suzuki et al., 2004; Chhatwal et al., 2005; Lafenêtre et al., 2007; Lutz, 2007; Niyuhire et al., 2007). We have recently shown that microinjecting the antagonist AM251 (6 ng) into the BLA or the CA1 significantly impairs extinction of inhibitory avoidance (Ganon-Elazar and Akirav, 2009; Abush and Akirav, 2010). Several studies suggest that the eCB system is not involved in the extinction of non-aversive memories (Hölter et al., 2005; Niyuhire et al., 2007).

On the other hand, studies have demonstrated that pharmacological activation of eCB signaling promotes extinction of fear memories. For example, Chhatwal et al. (2005) found that systemic administration of the eCB transporter AM404 (10 mg/kg) promotes extinction of fear that was conditioned using fear-potentiated

startle. This was replicated using systemic (Pamplona et al., 2008) and intracerebroventricular (Bitencourt et al., 2008) injections. In another study (Varvel et al., 2007), OL-135 (30 mg/kg), an inhibitor of FAAH, enhanced the rate of extinction in a water maze task. Pamplona et al. (2006) showed that WIN 55,212-2 (0.25 mg/kg) facilitates the extinction of contextual fear in the fear conditioning task and of spatial memory in the water maze reversal task. We have used the light–dark inhibitory avoidance procedure to demonstrate the effects of WIN 55,212-2 administered into the CA1 or the BLA on extinction. This procedure is dependent on both the amygdala and hippocampus as a single CS–US (context–footshock) pairing establishes a robust long-term memory, expressed as an increase in latency to enter the dark chamber at testing. Repeated retrieval of the avoidance response in the absence of the US induces extinction of inhibitory avoidance memory, meaning that the animal learns

that the context no longer predicts the footshock. We found that WIN 55,212-2 administered into the CA1 facilitates the extinction of inhibitory avoidance, with no effect on extinction kinetics when microinjected into the BLA (Ganon-Elazar and Akirav, 2009; Abush and Akirav, 2010).

Hence, the results of Marsicano et al. (2002) and subsequent investigations demonstrate that inhibition of eCB transmission robustly inhibits (or prolongs) fear extinction (Suzuki et al., 2004; Pamplona et al., 2006; Ganon-Elazar and Akirav, 2009; Abush and Akirav, 2010). Conversely, stimulation of eCB transmission accelerates fear extinction (Suzuki et al., 2004; Chhatwal et al., 2005; Barad et al., 2006; Abush and Akirav, 2010).

### COMPARING THE EFFECTS OF CANNABINOID AGONISTS ON AVERSIVE AND NON-AVERSIVE TASKS

It has been suggested that the neural processes underlying emotional memory formation (such as extinction learning) and non-emotional memories (such as spatial learning) are differentially sensitive to cannabinoid receptor activation (Chhatwal and Ressler, 2007). An intriguing question is whether cannabinoids have a similar effect on other types of emotional memories that do not involve fear and extinction learning.

We have recent findings suggesting that cannabinoid receptor activation has differential effects on learning and memory that are task-, brain region-, and memory stage-dependent (Segev and Akirav, 2011). We examined the effects of WIN 55,212-2 microinjected into the amygdala and the subiculum on the acquisition and retrieval of a neutral learning task (i.e., social discrimination) and an aversive learning task (i.e., contextual fear conditioning). The subiculum is the principal target of CA1 pyramidal cells. It functions as a mediator of hippocampal–cortical interaction and has been proposed to play an important role in the encoding and retrieval of long-term memory. In fear conditioning paradigms, the BLA plays a central role in the formation and consolidation of fear-related memory traces (LeDoux, 2003; Maren and Quirk, 2004), whereas the hippocampus's role is to integrate the features of the context and not to form a context–shock association (Fanselow, 1998). Unlike the aversive fear conditioning task, social discrimination is considered neutral or even rewarding. This finding was established using both conditioned place preference paradigms and T-maze learning rewarded by social interaction (Van den Berg et al., 1999). Social recognition processes depend on brain regions such as the medial amygdala, which modulates the initial social encounter and formation of social memory (Ferguson et al., 2001; Bielsky and Young, 2004) and the ventral hippocampus (Van Wimersma Greidanus and Maigret, 1996; Kogan et al., 2000).

We found that in the aversive contextual fear task, WIN 55,212-2 administered into the BLA impairs fear acquisition/consolidation, but not retrieval, whereas in the ventral subiculum (vSub), WIN 55,212-2 impairs fear retrieval. In the non-aversive or rewarding social discrimination task, WIN 55,212-2 into the vSub impairs acquisition/consolidation and retrieval, whereas in the medial amygdala, WIN 55,212-2 impairs acquisition (Segev and Akirav, 2011). These findings suggest that cannabinoid agonists can impair emotional (or aversive) as well as neutral (or rewarding) memory-related processes in a task-, region-, and memory stage-dependent manner. This is consistent with other

studies suggesting that exogenous acute cannabinoid treatment may have different outcomes depending on task aversiveness and the brain region involved (Suzuki et al., 2004; de Oliveira Alvares et al., 2005; Varvel et al., 2005; Ganon-Elazar and Akirav, 2009; Abush and Akirav, 2010).

### EFFECTS OF CANNABINOIDS ON STRESS AND ANXIETY

Considerable evidence suggests that cannabinoids are anxiolytics and modulate the behavioral and physiological response to stressful events (Viveros et al., 2007; Hill et al., 2010). Consequently, the effects of CB<sub>1</sub> agonists on learning and memory may be attributable to a general modulation of anxiety or stress levels and not to memory *per se*.

Stress is most readily defined as any stimulus that presents a challenge to homeostasis including any actual or potential disturbance of an individual's environment. The stress response enables the animal to adapt to the changing environment (Joëls and Baram, 2009). Fear is an adaptive component of the acute stress response to potentially dangerous stimuli that threaten the integrity of the individual. However, when disproportionate in its intensity, chronic, irreversible, and/or not associated with any actual risk, it constitutes a maladaptive response and may be symptomatic of anxiety-related neuropsychiatric disorders (Taber and Hurley, 2009).

Anxiety disorders are marked by excessive fear (and avoidance), often in response to specific objects or situations, in the absence of true danger, and they are common in the general population (Shin and Liberzon, 2010). As excessive fear is a key component of anxiety disorders, the search for the neurocircuitry of anxiety disorders has focused extensively on studies of fear circuits in animal models. These studies examined the neurocircuitry associated with fear responses in rats and mice using fear conditioning paradigms, inhibitory avoidance, and fear-potentiated startle models. The amygdala, PFC, and hippocampus have arisen as clear regions of interest in studies of anxiety disorders and are implicated in PTSD (Shin and Liberzon, 2010).

The hippocampus is often implicated in the neurobiology of stress. Mineralocorticoid and glucocorticoid receptors are expressed in high numbers within the hippocampus. Although stress-induced corticosteroid signaling in the hippocampus has a beneficial role in regulating the time course of the hypothalamic–pituitary–adrenal (HPA) axis stress response (De Kloet et al., 2005), prolonged glucocorticoid signaling can damage the hippocampus as measured by dendritic atrophy, decreased neurogenesis, and deficits in synaptic plasticity (McEwen and Gould, 1990; Sapolsky, 1996; McEwen, 1999; Meaney, 2001). In PTSD and major depression patients, hippocampus volumes are reduced (Bremner et al., 1995; Sheline et al., 1999; Woon and Hedges, 2008), and smaller hippocampal volumes are predictive of vulnerability to developing stress-related disorders (Pitman et al., 2006).

### ROLE OF THE ENDOCANNABINOID SYSTEM IN UNCONDITIONED STRESS AND ANXIETY

Results from many studies indicate that the eCB system modulates unconditioned stress- and anxiety-like responses (Viveros et al., 2005; Gorzalka et al., 2008; Lutz, 2009). A general conclusion that can be tentatively derived from the complicated and often contradictory literature is that inhibition of eCB signaling increases stress

and anxiety, while moderate increases in eCB signaling decrease stress and anxiety (Lutz, 2009; summarized in **Table 2**). The term “moderate” is used because strong stimulation of eCB signaling by high doses of CB<sub>1</sub> receptor agonists potentiates stress- and anxiety-like responses (Rodriguez de Fonseca et al., 1996; Scherma et al., 2008; Lutz, 2009). This biphasic effect has been demonstrated in animal models of anxiety (Lafenêtre et al., 2007; Hill and Gorzalka, 2009), and also in humans. Cannabis may induce aversive states in some smokers, precipitating anxiety and panic attacks (Hall and Solowij, 1998). Furthermore, THC administration may result in psychotic-like states (Linszen and van Amelsvoort, 2007). These bidirectional effects of cannabinoids observed in humans can be mimicked in laboratory animals. Hence, in models predictive of anxiolytic-like activity, low doses of CB<sub>1</sub> agonists tend to be anxiolytic and high doses tend to increase aversion and anxiety-related behaviors (Viveros et al., 2005).

Procedures used in studies on the role of eCBs in stress and anxiety evaluate the anxiolytic/anxiogenic effects of drugs by using standard tasks such as the elevated plus maze (EPM), social interaction, and defensive burying (Viveros et al., 2005; Lutz, 2009). Using the EPM, Patel and Hillard (2006) found that cannabinoid receptor agonists WIN 55212-2 (0.3–10 mg/kg) and CP55940 (0.001–0.3 mg/kg) administered systemically increase the time mice spend on the open arms (i.e., elicit an anxiolytic response) only at low doses. At the highest doses, both compounds alter overall locomotor activity. In contrast, THC (0.25–10 mg/kg) produces a dose-dependent reduction in time spent on open arms. The eCB uptake/catabolism inhibitor AM404 (0.3–10 mg/kg) produces an increase in time spent on the open arms at low doses and has no effect at the highest dose tested. The FAAH inhibitor URB597 (0.03–0.3 mg/kg) produces a monophasic, dose-dependent increase in time spent on the open arms. Systemic administration of the CB<sub>1</sub> receptor antagonists SR141716 (1–10 mg/kg) and AM251 (1–10 mg/kg) produce dose-related decreases in

time spent on open arms. Onaivi et al. (1990) have shown that THC induces increased aversion to the open arms of the EPM in both rats and mice that is similar to the aversion produced by anxiogenic agents. In contrast, mice treated with the agonists cannabidiol and nabilone spend a greater amount of time in the open arms of the maze, an effect similar to that produced by diazepam, the reference anxiolytic agent.

In the light–dark box, Berrendero and Maldonado (2002) have shown that the systemic administration of a low dose of THC (0.3 mg/kg) produces clear anxiolytic-like responses. The CB<sub>1</sub> cannabinoid receptor antagonist SR 141716A (0.5 mg/kg) completely blocks the anxiolytic-like response induced by THC, suggesting that this effect is mediated by CB<sub>1</sub> cannabinoid receptors. In another study, systemic administration of the FAAH inhibitors URB597 and URB532 reduces anxiety-related behavior in the rat elevated zero-maze and in isolation-induced ultrasonic vocalization tests (Kathuria et al., 2003). These effects are dose-dependent and blocked by the antagonist rimonabant. The FAAH inhibitor and eCB re-uptake inhibitor AM404 also exhibit a dose-dependent anxiolytic profile in the EPM, defensive withdrawal test, and ultrasonic vocalization test (Bortolato et al., 2006). URB597 has also been shown to be anxiolytic in the rat EPM and open-field tests (Hill et al., 2007) and has recently been shown to reduce anxiety-related behavior in the EPM in Syrian hamsters (Moise et al., 2008).

Ribeiro et al. (2009) examined the dose-response effects of exogenous anandamide at doses of 0.01, 0.1, and 1.0 mg/kg in mice sequentially submitted to the open field and EPM. Systemically administered at 0.1 mg/kg (but not at 0.01 or 1 mg/kg), anandamide increases the time spent and the distance covered in the central zone of the open field, as well as exploration of the open arms of the EPM. Recently, Rubino et al. (2008b) demonstrated that the anxiolytic-like effect of a low anandamide dose is reversed by administration of the antagonist AM251, whereas the anxiogenic-like effect is

**Table 2 | Effects of cannabinoids on anxiety-related responses.**

Agonist	Species	Doses	Apparatus	Effects	References
WIN 55,212-2	Mice	0.3–10 mg/kg	EPM	+	Patel and Hillard (2006)
CP55940	Mice	0.001–0.3 mg/kg	EPM	+	Patel and Hillard (2006)
THC	Mice	0.25–10 mg/kg	EPM	–	Patel and Hillard (2006)
	Rats	1–10 mg/kg	EPM	–	Onaivi et al. (1990)
	Mice	10–20 mg/kg	EPM	–	Onaivi et al. (1990)
	Mice	0.3 mg/kg	Light–dark box	+	Berrendero and Maldonado (2002)
AM404	Mice	0.3–10 mg/kg	EPM	+	Patel and Hillard (2006)
URB597	Mice	0.03–0.3 mg/kg	EPM	+	Patel and Hillard (2006)
	Rats	0.05–0.1 mg/kg	Zero-maze	+	Kathuria et al. (2003)
URB532	Rats	0.1–10 mg/kg	Ultrasonic test	+	Kathuria et al. (2003)
			Ultrasonic test	+	Kathuria et al. (2003)
Nabilone	Mice	10–100 µg/kg	EPM	+	Onaivi et al. (1990)
Cannabidiol	Mice	1–10 mg/kg	EPM	+	Onaivi et al. (1990)
Anandamide	Mice	0.1 mg/kg	EPM	+	Ribeiro et al. (2009)
			Open field	+	

Effects: –, anxiogenic effect; +, anxiolytic effect. EPM, elevated plus maze.

inhibited by pre-treatment with capsazepine, a transient receptor potential vanilloid type 1 (TRPV1) receptor antagonist. The authors suggested that the anxiolytic effect evoked by anandamide might be due to the interaction with the CB<sub>1</sub> cannabinoid receptor, whereas vanilloid receptors seem to be involved in the anxiogenic action of anandamide (Rubino et al., 2008b). Marsch et al. (2007) reported that TRPV1 “null” mice exhibit a significantly reduced response to anxiogenic stimuli. Therefore, the anandamide-induced inverted U-shape pattern might be based on the fact that the intrinsic efficacy of anandamide on TRPV1 is relatively low compared to that observed on the CB<sub>1</sub> receptor (Ross, 2003).

Transgenic mice deficient for FAAH, the enzyme that degrades anandamide, demonstrate reduced anxiety-like behavior in the EPM and light–dark box compared with wild-type mice and these effects are prevented by systemic administration of the antagonist rimonabant (Moreira et al., 2008). By contrast, transgenic mice lacking expression of the CB<sub>1</sub> receptor demonstrate an anxiogenic profile in the EPM, the light–dark box, open-field arena, and social interaction test (Haller et al., 2002, 2004; Maccarrone et al., 2002; Martin et al., 2002; Urigüen et al., 2004) and demonstrate impaired stress coping behavior in the forced swim test (Steiner et al., 2008). Similarly, CB<sub>1</sub> receptor antagonists increase anxiety-related behaviors in the EPM (Patel and Hillard, 2006). Taken together, these studies suggest that eCBs act at CB<sub>1</sub> receptors to reduce anxiety.

#### ROLE OF THE ENDOCANNABINOID SYSTEM IN CONDITIONED FEAR AND ANXIETY

Understanding the role of the eCB system in conditioned fear and aversive memories is important because a number of anxiety disorders, including PTSD and phobias, are thought to result from dysregulated fear neurocircuitry (Rauch et al., 2006). Investigators have examined the effect of CB<sub>1</sub> receptor agonists and antagonists on contextual and cue fear conditioning. Results from these studies were somewhat mixed. In rats, systemic injections of the CB<sub>1</sub> receptor antagonist AM251 enhance both the acquisition and expression of cue fear conditioning (Arenos et al., 2006; Reich et al., 2008). Administering AM251 (5 mg/kg, i.p) during tone–footshock conditioning enhances acquisition of freezing behavior for both trace fear conditioning (hippocampal-dependent) and delay fear conditioning (amygdala-dependent; Reich et al., 2008). Recently, we used an inhibitory avoidance task and found that microinjecting AM251 (6 ng) into the BLA significantly enhances conditioned avoidance but has no effect on conditioning when microinjected into the hippocampal CA1 area (Ganon-Elazar and Akirav, 2009; Abush and Akirav, 2010). However, others have shown that mice lacking the CB<sub>1</sub> receptor or systemically administered with the CB<sub>1</sub> receptor antagonist AM251 (0.3–3 mg/kg) 30 min before behavioral testing show no contextually induced fear response (Mikics et al., 2006). Furthermore, the CB<sub>1</sub> receptor antagonist rimonabant or genetic deletion of the CB<sub>1</sub> receptor has no effect on the acquisition of cue and context fear conditioning in mice (Marsicano et al., 2002; Suzuki et al., 2004). On the other hand, cue-fear-potentiated startle is decreased by medial PFC injections of the CB<sub>1</sub> receptor agonist WIN 55212-2 or the FAAH inhibitor URB597 (Lin et al., 2008, 2009) and contextual fear conditioning is decreased by dorsolateral periaqueductal gray injections of either anandamide or

the anandamide transport inhibitor AM404 (Resstel et al., 2008). Overall it appears that, as in the case of unconditioned fear, inhibition of eCB transmission increases fear while moderate stimulation of eCB transmission decreases fear.

#### THE INVOLVEMENT OF THE HIPPOCAMPUS IN ENDOCANNABINOID MODULATION OF STRESS AND ANXIETY

Techniques based on intracranial injections of cannabinoids in rats revealed that activation of CB<sub>1</sub> receptors is involved in inducing anxiolytic- or antidepressant-like effects (Bambico et al., 2007; Moreira et al., 2007; Rubino et al., 2008a,b). For example, Rubino et al. (2008a) found that low doses of THC microinjected into the PFC (10 µg) or ventral hippocampus (5 µg) in rats induces an anxiolytic-like response during tests in the EPM, while higher doses do not show an anxiolytic effect and even seem to switch into an anxiogenic profile. Nevertheless, other studies demonstrated that eCB activation in the amygdala and dorsal hippocampus results in an anxiogenic-like response. Low THC doses (1 µg) in the BLA produce an anxiogenic-like response whereas higher doses are ineffective (Rubino et al., 2008a). WIN-55212-2 in the dorsal hippocampus (2.5 and 5 µg) produces a significant anxiogenic-like effect in rats that is reversed by AM251 (Roohbakhsh et al., 2007).

Local infusion of cannabinoid compounds into specific brain areas might be instrumental in identifying neural pathways and neuroanatomically separated CB<sub>1</sub> receptor subpopulations that may play distinct roles in and mediate the opposing actions of cannabinoids, notably, anxiolytic versus anxiogenic effects (Moreira et al., 2007; Viveros et al., 2007). We examined the role of cannabinoids in modulating aversive and non-aversive learning paradigms in the hippocampus and amygdala (Ganon-Elazar and Akirav, 2009; Abush and Akirav, 2010; Segev and Akirav, 2011). Microinjecting the antagonist AM251 (6 ng) or the agonist WIN-55212-2 (5 µg) into the BLA, CA1, or vSub had no effect on anxiety levels as measured in the open-field, pain sensitivity (Ganon-Elazar and Akirav, 2009; Abush and Akirav, 2010; Segev and Akirav, 2011), or EPM tests (Abush and Akirav, 2010). However, both agonist and antagonist had profound effects on aversive and non-aversive learning tasks. These findings suggest that in these studies the impairing and facilitating effects of local infusions of WIN-55212-2 on learning and memory are probably not attributable to a general modulation of anxiety. Nevertheless, the effects of cannabinoids on the interplay between anxiety and memory processes are difficult to separate and further examination of the effects of different cannabinoids is required.

To summarize the role of the eCB system in stress, anxiety, and conditioned fear, there is a general consensus that the effects of cannabinoid agonists on anxiety seem to be biphasic, with low doses being anxiolytic and high doses being ineffective or possibly anxiogenic. There are several important characteristics of the eCB system that might explain these different effects of eCB modulation. First, in a physiological situation, eCB synthesis, and thus CB<sub>1</sub> receptor activation, occurs in particular activated neuronal circuits. This is a notable difference from the situation following pharmacological treatment with receptor agonists, when the agent activates all CB<sub>1</sub> receptors in the brain regardless of their specific involvement in a particular physiological process. Second, the CB<sub>1</sub>

receptor is expressed in diverse brain structures of relevance to psychiatric disorders and is mainly located presynaptically where it can suppress the release of other neurotransmitters (Marsicano and Lutz, 1999, 2006; Mackie, 2005). These neurotransmitters include the main inhibitory neurotransmitter GABA, the main excitatory neurotransmitter glutamate, as well as acetylcholine, noradrenaline, and serotonin (Katona et al., 1999; Harkany et al., 2005; Monory et al., 2006; Häring et al., 2007; Oropeza et al., 2007). Thus, synthetic compounds delivered systemically lack both the spatial and temporal specificity of endogenous compounds (Lafenêtre et al., 2007; Viveros et al., 2007; Moreira and Lutz, 2008). This may explain not only the bell-shaped relationship between dose and effect that some studies have observed, but also why elevation of eCB levels sometimes has effects that are different from those observed with exogenous cannabinoids. Finally, the diversity of eCB ligands with their multiple synthetic and degradation pathways adds a further level of complexity to the eCB system (Di Marzo, 2008).

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